



GTI Diagnostic Clinic Submission Form

OFFICE USE ONLY

Service summary: _____ SAMPLE ID: _____

Total charges: _____ Receipt #: _____

CLIENT INFORMATION

Primary Contact: _____ Alternative contact: _____

Email: _____ Phone: _____

Cell: _____ Fax: _____

Business Name: _____

Address: _____ City: _____ Prov: _____ Postal code: _____

THIRD PARTY PAYMENT INFO

Name: _____ email: _____

IMPORTANT: PLEASE FILL IN ALL CONTACT INFORMATION AND BACKGROUND DETAILS FOR SAMPLE. THIS WILL HELP ENSURE A QUICK AND ACCURATE DIAGNOSIS. SEND VIA COURIER TO ENSURE NEXT DAY ARRIVAL. INSTRUCTIONS FOR TAKING A SAMPLE ARE LOCATED ON THE BACK OF THIS FORM

Send Sample to:

The GTI Diagnostic Clinic c/o Dr. Katerina S. Jordan
Dept of Plant Agriculture, E. C. Bovey Building
University of Guelph, 50 Stone Road East
Guelph, ON N1G 2W1

Drop off:

4224 E.C.Bovey Building (park at meters on South Ring Rd.)
University of Guelph (intersection of Gordon St. and South Ring Rd.)
Contact clinic to make appointment or leave on table outside lab

Service option: Diagnosis via e-mail or telephone
Per sample \$120.00

Nematode extraction only
Per sample \$50.00

Payment Information: Payment will be processed once sample has been processed. Info will not be kept on file

Cheque (enclosed with sample): Payable to "University of Guelph"

VISA / MC #: _____ Expiry ____ / ____

Name on card: _____ SIGNATURE: _____



Contact Information:
Phone: 519-824-4120 x 58873
Email: turfdiag@uoguelph.ca
Fax: 519-766-1704

SAMPLE INFORMATION SHEET

Date symptoms first appeared*: _____ Date sample taken*: _____

Weather conditions at onset of symptoms (Temperature, rainfall, other)*: _____

Species (if known): _____ Cultivar/Variety (if known): _____

Check the following as they pertain to your problem (information with asterisks will likely lead to a faster diagnosis)

LOCATION*	ORIGIN/AGE*	PATTERN OF DAMAGE	DEGREE OF INJURY	SOIL CONDITIONS
<input type="checkbox"/> Green#	<input type="checkbox"/> Sod	<input type="checkbox"/> General	<input type="checkbox"/> Light	pH* _____
<input type="checkbox"/> Tee #	<input type="checkbox"/> Seeded	<input type="checkbox"/> Scattered	<input type="checkbox"/> Moderate	Thatch levels:
<input type="checkbox"/> Fairway#	<input type="checkbox"/> Thinning	<input type="checkbox"/> Severe	SITE CONDITIONS	Low _____
<input type="checkbox"/> Lawn	Age* : _____	<input type="checkbox"/> Rings	<input type="checkbox"/> Compacted	Moderate _____
<input type="checkbox"/> Sports field		<input type="checkbox"/> Patches	<input type="checkbox"/> Heavy traffic	High _____
<input type="checkbox"/> Sod Farm		<input type="checkbox"/> Leaf spots		Root zone material _____

Describe the problem in detail (i.e. symptoms, plant parts affected, distribution of symptoms):

Disease or disorder history of the site:

Were fungicides or fertilizers applied recently? Specify type of products (s) and date (s) of application:

Additional comments and specific requests:

Photos are also helpful. Please include photos or send via email to turfdiag@uoguelph.ca. Be sure to include your name and other information so we know which picture is for which sample.

Instructions for Taking and Submitting a Sample

DISEASE/PATHOLOGY SAMPLING

1. Sample BEFORE you treat with fungicides. Fungicide application destroys signs of pathogens impeding diagnosis.
2. Sample should be 10 to 15 cm² (cup cutter size is ideal) and include foliage, thatch and at least 5 cm of roots and soil
3. Sample should show a range of symptoms and include healthy, slightly affected and severely affected grass. Take the sample from the outside edge of a ring or patch that includes healthy and unhealthy turf. If symptoms are general, collect the sample from an area with intermediate severity. NOTE: a completely dead sample is not suitable for diagnosis.
4. Do not allow the sample to dry out or to be exposed to excessive heat or cold prior to submission.
5. Sample should be wrapped in newspaper and then in plastic and placed in a sturdy box.

NEMATODE SAMPLING

- A cup cutter can be used but will limit total sampling area and can be expensive for shipping; It is better to take multiple small extraction plugs (2cm diameter minimum) from various areas throughout affected zones
- 10 cores is the minimum – 20 to 30 is better
- Cores should be 10 cm or 4" deep
- The top 2 cm of grass, thatch and plant material can be removed from the plugs/cores

METHOD 1:

1. Take samples only from the edge of affected areas – DO NOT include dead patches or asymptomatic areas.
2. This method relies on threshold data to confirm whether the issue you are experiencing is related to nematodes or not.
3. This method does not take into account baseline populations in asymptomatic areas, and therefore produces limited information regarding nematodes on your property.

METHOD 2: (Recommended)

1. Take separate samples from both symptomatic areas and asymptomatic areas and request that each sample be assessed separately.
2. Keep these samples separate from each other as they will be considered two separate samples.
3. This method is more expensive but will produce information on your baseline levels in asymptomatic areas as well as threshold population data in the affected areas.

METHOD 3:

1. Take samples systematically throughout entire area (ie. every 15 feet) including both affected and non-affected areas and combine the cores into one sample.
2. This method is useful for determining overall population levels but is not useful in associating symptoms with nematode populations.

WHEN SUBMITTING A SAMPLE:

1. Fill out the submission form as completely as possible and include it with the sample
2. Do not send samples by regular mail or over a weekend! They must be sent so that they arrive **NEXT DAY**.
3. **Photos are not necessary but do help with the diagnostic process.**

You do not need to include this sheet with your submission