



GTI Diagnostic Clinic Submission Form

OFFICE USE ONLY

Service summary: \_\_\_\_\_ SAMPLE ID: \_\_\_\_\_
Total charges: \_\_\_\_\_ Receipt #: \_\_\_\_\_

CLIENT INFORMATION

Primary Contact: \_\_\_\_\_ Alternative contact: \_\_\_\_\_
Email: \_\_\_\_\_ Phone: \_\_\_\_\_
Cell: \_\_\_\_\_
Business Name: \_\_\_\_\_
Address: \_\_\_\_\_ City: \_\_\_\_\_ Prov: \_\_\_\_\_ Postal code: \_\_\_\_\_

THIRD PARTY PAYMENT INFO

Name: \_\_\_\_\_ email: \_\_\_\_\_

IMPORTANT: ALL SAMPLES SHOULD BE SENT VIA COURIER TO ENSURE NEXT DAY ARRIVAL. ALL PACKAGES SHOULD BE MARKED AS PERISHABLE SO THEY ARE HANDLED WITH CARE AND REACH THE LAB ASAP. INSTRUCTIONS FOR TAKING SAMPLES ARE LOCATED ON THE BACK OF THIS FORM.

Send Sample to: The GTI Diagnostic Clinic c/o Dr. Katerina S. Jordan Dept of Plant Agriculture, E. C. Bovey Building University of Guelph, 50 Stone Road East Guelph, ON N1G 2W1

Drop off: Please email (turfdiag@uoguelph.ca) or text Dr. Katerina Jordan (519-241-2566) to schedule a drop-off.

- Service option: [ ] Diagnosis via e-mail or telephone Per sample \$135.00
[ ] Nematode extraction only Per sample \$65.00

Payment Information: Payment will be processed once sample has been processed. Info will not be kept on file

[ ] Cheque (enclosed with sample): Payable to "University of Guelph"

[ ] VISA / MC #: \_\_\_\_\_ Expiry \_\_\_\_ / \_\_\_\_

Name on card: \_\_\_\_\_ SIGNATURE: \_\_\_\_\_



**Contact Information:**  
Phone: 519-824-4120 x 58873  
Email: [turfdiag@uoguelph.ca](mailto:turfdiag@uoguelph.ca)

## SAMPLE INFORMATION SHEET

Date symptoms first appeared\*: \_\_\_\_\_ Date sample taken\*: \_\_\_\_\_

Weather conditions at onset of symptoms (Temperature, rainfall, other)\*: \_\_\_\_\_

Species (if known): \_\_\_\_\_ Cultivar/Variety (if known): \_\_\_\_\_

Check the following as they pertain to your problem (information with asterisks will likely lead to a faster diagnosis)

LOCATION*	ORIGIN/AGE*	PATTERN OF DAMAGE	DEGREE OF INJURY	SOIL CONDITIONS
<input type="checkbox"/> Green#	<input type="checkbox"/> Sod	<input type="checkbox"/> General	<input type="checkbox"/> Light	<b>pH*</b> _____
<input type="checkbox"/> Tee #	<input type="checkbox"/> Seeded	<input type="checkbox"/> Scattered	<input type="checkbox"/> Moderate	Thatch levels:
<input type="checkbox"/> Fairway#	<input type="checkbox"/> Thinning	<input type="checkbox"/> Severe	<b>SITE CONDITIONS</b>	Low _____
<input type="checkbox"/> Lawn	<b>Age*</b> : _____	<input type="checkbox"/> Rings	<input type="checkbox"/> Compacted	Moderate _____
<input type="checkbox"/> Sports field		<input type="checkbox"/> Patches	<input type="checkbox"/> Heavy traffic	High _____
<input type="checkbox"/> Sod Farm		<input type="checkbox"/> Leaf spots		Root zone material _____

Describe the problem in detail (i.e. symptoms, plant parts affected, distribution of symptoms):

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Disease or disorder history of the site:

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Were fungicides or fertilizers applied recently? Specify type of products (s) and date (s) of application:

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Additional comments and specific requests:

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***PHOTOS ARE EXTREMELY HELPFUL! Please include printed photos with your mailed sample or send via email to [turfdiag@uoguelph.ca](mailto:turfdiag@uoguelph.ca). Be sure to include your name and other information so we can correlate the right picture with the right sample.***

## Instructions for Taking and Submitting a Sample

### DISEASE/PATHOLOGY SAMPLING

1. Sample BEFORE you treat with fungicides. Fungicide application destroys signs of pathogens impeding diagnosis.
2. Sample should be 10 to 15 cm<sup>2</sup> (cup cutter size is ideal) and include foliage, thatch and at least 5 cm of roots and soil
3. Sample should show a range of symptoms and include healthy, slightly affected and severely affected grass. Take the sample from the outside edge of a ring or patch that includes healthy and unhealthy turf. If symptoms are general, collect the sample from an area with intermediate severity. NOTE: a completely dead sample is not suitable for diagnosis.
4. Do not allow the sample to dry out or to be exposed to excessive heat or cold prior to submission.
5. Sample should be wrapped in newspaper and then in plastic and placed in a sturdy box.

### NEMATODE SAMPLING

- A cup cutter can be used but will limit total sampling area and can be expensive for shipping; It is better to take multiple small extraction plugs (2cm diameter minimum) from various areas throughout affected zones
- 10 cores is the minimum – 20 to 30 is better
- Cores should be 10 cm or 4" deep
- The top 2 cm of grass, thatch and plant material can be removed from the plugs/cores

#### METHOD 1:

1. Take samples only from the edge of affected areas – DO NOT include dead patches or asymptomatic areas.
2. This method relies on threshold data to confirm whether the issue you are experiencing is related to nematodes or not.
3. This method does not take into account baseline populations in asymptomatic areas, and therefore produces limited information regarding nematodes on your property.

#### METHOD 2: (Recommended)

1. Take separate samples from both symptomatic areas and asymptomatic areas and request that each sample be assessed separately.
2. Keep these samples separate from each other as they will be considered two separate samples.
3. This method is more expensive but will produce information on your baseline levels in asymptomatic areas as well as threshold population data in the affected areas.

#### METHOD 3:

1. Take samples systematically throughout entire area (ie. every 15 feet) including both affected and non-affected areas and combine the cores into one sample.
2. This method is useful for determining overall population levels but is not useful in associating symptoms with nematode populations.

### WHEN SUBMITTING A SAMPLE:

1. Fill out the submission form as completely as possible and include it with the sample
2. Do not send samples by regular mail or over a weekend! They must be sent so that they arrive **NEXT DAY**.
3. **Photos are not necessary but do help with the diagnostic process.**

*\*You do not need to include this sheet with your submission\**